

Effect of Doxycycline on Atherosclerosis: From Bench to Bedside

Gastón A. Rodriguez-Granillo^{1,2,3#,*} Agustina Rodriguez-Granillo^{4,5#}, and José Milei¹

¹*Instituto de Investigaciones Cardiológicas. UBA-CONICET, Buenos Aires, Argentina,* ²*Department of Cardiovascular Imaging, Otamendi Hospital, Buenos Aires, Argentina,* ³*Department of Cardiovascular Imaging, Clinica La Sagrada Familia, Buenos Aires, Argentina,* ⁴*Center for Advanced Biotechnology and Medicine, University of Medicine and Dentistry of New Jersey, NJ, USA,* ⁵*Cardiovascular Research Center (CECI), Buenos Aires, Argentina*

Received: June 30, 2010; Accepted: September 7, 2010; Revised: September 7, 2010

Abstract: Matrix metalloproteinases (MMPs) have a pivotal role in the natural history of atherosclerosis and its cardiovascular consequences. Non-selective MMP inhibition with doxycycline appears as a potential strategy to reduce the residual risk observed in patients already at intensive lipid lowering strategies. However, specific MMPs have different and even contradicting roles in the natural history of atherosclerosis, rendering broad spectrum MMP inhibition an important yet somewhat simplistic approach towards residual risk reduction in coronary atherosclerosis. Overall, the balance of non-selective MMP inhibition might shift to the favorable side in particular settings such as in acute coronary syndromes, where in addition to its potential plaque stabilization properties, doxycycline shows promise in preventing ischemia-reperfusion injury and left ventricular remodeling. Nevertheless, to date, most animal models used do not represent advanced coronary atherosclerosis seen in humans, and large and well-designed clinical studies are lacking. We discuss the available evidence and recent patents supporting the role of doxycycline in atherosclerosis.

Keywords: Inflammation, coronary, metalloproteinase, remodeling, atherothrombosis, MMP inhibitor, apo E deficient mice.

INTRODUCTION

Cardiovascular disease is the main cause of morbidity and mortality in the Western Hemisphere, accounts for > 500,000 deaths each year in the US alone, and doubles the mortality attributed to cancer [1]. Histopathological studies have established that atherosclerotic plaque composition as well as coronary artery remodeling patterns have a pivotal role in the etiology of acute coronary thrombosis, independently of the underlying stenosis [2].

Several systemic strategies have demonstrated their effectiveness in primary and secondary prevention of coronary artery disease (CAD). Among them, statins have shown a consistent decline in low density lipoprotein cholesterol (LDL-C) levels between 25% and 35%, with a significant reduction in the relative risk of myocardial infarction (MI) and death ranging between 29% and 35% [3-5]. Nevertheless, in spite of significant improvement in prevention, diagnosis and treatment of cardiovascular disease, sudden cardiac death or unheralded acute coronary syndromes (ACS) remain common initial manifestations of coronary atherosclerosis [6, 7]. These events are mainly attributed to a significant residual risk observed in approximately 70% of patients under optimal anti-atherosclerotic therapies with statins, angiotensin-converting enzyme inhibitors and aspirin, among others. Although, implementation of

aggressive lipid lowering therapies with target LDL-C levels < 70mg/dl in high risk patients have demonstrated an additional risk reduction of 16%, even in these cases a 22% residual risk is observed [8]. Furthermore, a recent pooled analysis of 7 serial intravascular ultrasound trials including 3437 patients with CAD demonstrated that despite achieving a LDL-C < 70mg/dl level, more than 20% of patients continued to show plaque progression, high-lighting the multifactorial nature of atherosclerosis and the need for improvement in primary and secondary prevention strategies, with the incorporation of alternative drugs that contribute to reduce the significant residual risk [9].

Matrix metalloproteinases (MMPs) are a family of endopeptidases that act as regulators of the extracellular matrix (ECM), playing an essential role in the evolution of inflammatory processes and hence, in the natural history of atherosclerosis [10]. We therefore review the available evidence about the effect of a broad-spectrum MMP inhibitor (MMPI), doxycycline, on atherosclerosis.

MMPs AS VALID BUT COMPLEX CLINICAL TARGETS

Human MMPs are a family of at least 23 endopeptidases (although previously considered to be more, 23 human MMPs are currently identified by the Universal Protein Resource, www.uniprot.org) involved in the remodeling of several components of the ECM [11-14]. They participate in almost every biological process involving ECM remodeling, such as angiogenesis, embryogenesis, tissue remodeling and wound healing [15]. Although it was originally believed that the role of MMPs was essentially to degrade the ECM, it is

*Address correspondence to this author at the Department of Cardiovascular Imaging, Otamendi Hospital, Azcuena 870. C1115AAB. Buenos Aires, Argentina; Tel/Fax: +5411 49648740; E-mail: grodriguezgranillo@gmail.com

now well-known that the function of MMPs is far more complex [13]. In fact, it has been shown that MMPs can act on as many non-ECM substrates as ECM substrates [16]. MMP substrates include ECM molecules and regulators, chemokines, cytokines, growth factors, angiogenic factors, receptors, proteases, metabolic enzymes, and proteins involved in cell adhesion and motility [13]. Therefore, MMPs should be considered as cell-signaling regulators rather than as solely destructive proteases [17].

Under normal physiological conditions, MMP activity is tightly regulated at the transcriptional and post-translational levels, by zymogen activation and by endogenous inhibitors [15]. Tissue inhibitors of MMPs (TIMPs) are specific, potent and natural MMPIs that bind to these enzymes and block their activity. Disruption of this balance, which results in an overexpression of MMPs, has been associated with severe human pathologies including cardiovascular diseases, cancer, rheumatoid arthritis, neurological disorders and periodontitis. Thus, considerable efforts have been made to develop potent and selective MMPIs to treat these diseases [18-21]. However, after 30 years of extensive research, only doxycycline (Periostat[®], CollaGenex Pharmaceuticals) has been clinically approved as a broad-spectrum MMPI for the treatment of periodontal disease [22, 23]. Previous clinical trials with synthetic MMPIs were disappointing because of severe side effects and poor survival rates [12, 24-27]. This failure has been mainly due to the complex biology of the MMPs, the use of broad-spectrum MMPIs, and limitations in the design of the clinical trials [26, 27].

The structural redundancy but functional diversification among the different subclasses of MMPs is the main challenge when developing selective MMPIs. Although significant overlap in the substrates that MMPs can cleave *in vitro* [11], the efficiency, as also the expression patterns and turnover in a given tissue, can vary [28, 29]. Therefore, for a given pathology, some MMPs might act as drug targets and others as anti-targets [12], underscoring the importance of MMPI specificity.

THE STRUCTURAL AND FUNCTIONAL COMPLEXITY OF MMPs

MMPs are zinc (Zn)-dependant enzymes comprised of shared structural modules [30]. The interaction between these domains is crucial for specific substrate binding and processing [30]. Based on domain organization and substrate specificity, MMPs can be clustered into several groups Fig. (1). [31]. The archetypical domain arrangement consists of a N-terminal propeptide, a catalytic MMP domain, a linker region (hinge) and a hemopexin-like C-terminal domain. The catalytic domain contains the characteristic Zn-binding sequence with three conserved histidine (His) residues, which serve as the Zn-ligands, and one glutamic acid (Glu), which facilitates catalysis, and is stabilized by a structural Zn and up to three calcium (Ca) ions. The catalytic domains of all MMPs are essentially superimposable containing a shallow active-site cleft that binds a peptide-substrate [32]. Substrate binding is dictated by the structure of this active site, including a pocket called the S₁' pocket, a main determining factor for substrate specificity [31].

All MMPs are either secreted or anchored to the plasma membrane. A hallmark of the MMP family is its regulation by zymogen (pro-MMP) activation. Most MMPs exist in an inactive or latent form inside the cells, in which the prodomain interacts with the catalytic domain, blocking the active site. The enzymes are activated when this interaction is relieved, upon removal of the propeptide or a conformational change [33], once they are secreted. Exceptions are MMP-11 (stromelysin-3), -21, -23, -28 (epilysin) and the six membrane-type (MT)-MMPs, which are activated by furin in the endosomal pathway [15]. The extracellular activation of most MMPs can be initiated by other already active MMPs or by several serine proteinases [11].

MMPs also share a common reaction mechanism in which the peptidic substrate is cleaved. During the proposed proteolytic mechanism Fig. (2). the carbonyl group of the scissile peptide bond of the substrate is directed towards the catalytic Zn and becomes polarized [18]. The Zn-bound water molecule is activated by the catalytic Glu properly oriented to attack the electrophilic carbonyl carbon. The resulting tetrahedral intermediate is presumably stabilized by the Zn ion. Additionally, one water proton is shuttled via the Glu carboxylate to the amino group of the scissile bond. After the simultaneous break of the peptide bond and the transfer of another proton to the amino group, the two product fragments leave the active site.

NATURAL HISTORY OF ATHEROSCLEROSIS

Since atherosclerosis is primarily an inflammatory disease, MMPs play a pivotal role in the development and natural history of atherosclerotic plaques [34].

In human atherosclerotic plaques, the intima comprises a hyaluronan-rich matrix with sparse vascular smooth muscle cells (VSMC) [35]. Basement membranes contain type IV collagen, laminin, and heparan sulfate proteoglycans such as perlecan [36] and syndecans [37]. The media comprises contractile VSMC surrounded by a basement membrane [38], few macrophages and fibroblasts [39]. The medial interstitial matrix contains types I and III collagen, elastin, and a number of glycoproteins, namely fibronectin, vitronectin, tenascin, and thrombospondin, along with chondroitin/dermatan sulfate proteoglycans, such as versican [40]. Finally, the adventitia comprises fibroblasts and *vasa vasorum* within a loose interstitial matrix [41].

In response to a vascular insult or biochemical stimuli, intimal thickening occurs mediated by a variety of cells along with accumulation of new ECM [42]. Provided that the stimuli remains, uptake of LDL which will subsequently transform into oxidized LDL is followed by infiltration by circulating monocytes, which convert to macrophages and incorporate oxidized LDL to become foam cells [43]. Later, apoptosis of macrophages and dumping of their lipid contents results in the formation of a fibrous cap overlying a large lipid core [44]. The fibrous cap is originated by migration of contractile VSMC from the media, which explains the medial thinning commonly observed in atherosclerotic plaques [45].

It has been established that coronary plaque rupture, resulting in ACS, is the cause of death in a large proportion

Archetypal MMPs*Collagenases*

Collagenase-1 (MMP-1)

Collagenase-2 (MMP-8)

Collagenase-3 (MMP-13)

Stromelysins

Stromelysin-1 (MMP-3)

Stromelysin-2 (MMP-10)

Other MMPs

Metalloelastase (MMP-12)

MMP-19

Enamelysin (MMP-20)

MMP-27 (C-MMP)

**Matrilysins**

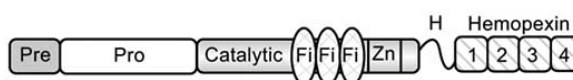
Matrilysin-1 (MMP-7)

Matrilysin-2 (MMP-26)

**Gelatinases**

Gelatinase-A (MMP-2)

Gelatinase-B (MMP-9)

**Furin-activated MMPs***Secreted*

Stromelysin-3 (MMP-11)

Epilysin (MMP-28)

*Vitronectin-like insert*

MMP-21 (XMMP)

*Membrane-associated**Transmembrane*

MT1-MMP (MMP-14)

MT2-MMP (MMP-15)

MT3-MMP (MMP-16)

MT5-MMP (MMP-24)

*GPI-anchored*

MT4-MMP (MMP-17)

MT6-MMP (MMP-25)

*Type-II transmembrane*

MMP-23



Fig. (1). Schematic representation of the structure of the 23 human MMPs. They are classified into four different groups on the basis of domain organization. Archetypal secreted MMPs contain a signal peptide (Pre), a propeptide (Pro), a catalytic domain that binds Zn, a linker (H), and a hemopexin C-terminal domain. Matrilysins contain the minimal domain organization that is required for function. Gelatinases incorporate three fibronectin (Fi) type II modules that improve collagen and gelatin degradation efficiency. Convertase-activatable MMPs contain a basic insert in the propeptide domain that is cleaved by furin-like proteases (Fu). This group includes the three secreted MMPs (MMP-11, MMP-21, MMP-28), the six membrane-type (MT)-MMPs and an unusual type-II transmembrane (TM) MMP (MMP-23). MMP21 contains a vitronectin-like (Vn) insert in the propeptide. MT-MMPs are inserted in the membrane by a type-I TM or GPI (glycosylphosphatidylinositol) anchor. Another linker (TML) connects these segments with the soluble archetypal core. MT-MMPs can also have a cytoplasmic (Cy) tail. MMP-23 contains a unique cysteine array (CA) and immunoglobulin (Ig)-like domains in its C-terminal region.

of sudden death patients [46]. Despite its pre-conceived dire prognosis, retrospective studies have determined that plaque rupture is a common finding in both coronary and non-coronary sudden death patients [46, 47]. In addition, clinically silent plaque rupture has been identified as a cause of plaque progression [48, 49]. The fate of a given atherosclerotic plaque is linked not only to its severity but

also to its histological composition, and the presence of a lipid-rich necrotic core has been consistently related to plaque fissuring [50, 51]. Plaque rupture typically occurs in regions of high mechanical stress and where collagen is depleted by matrix destruction, weakening the fibrous cap to the point where it can no longer resist the cyclical strain caused by the cardiac cycle [52, 53].

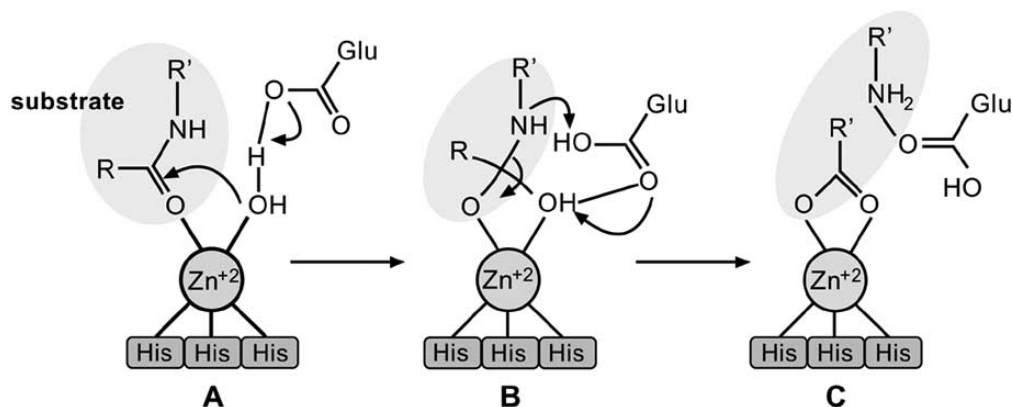


Fig. (2). Proposed reaction mechanism of peptide hydrolysis for MMPs. Within the MMP's catalytic domain, the catalytic Zn that is coordinated by three histidine (His) residues binds the carbonyl group of the scissile peptide bond of the substrate (A). The Zn-bound water molecule is activated by the catalytic glutamic acid (Glu), which is properly oriented to attack the electrophilic carbonyl carbon of the substrate, leading to a tetrahedral intermediate (B). Next, one water proton is shuttled via the Glu carboxylate to the amino group of the scissile bond. After the simultaneous break of the peptide bond and the transfer of another proton to the amino group (C), the two product fragments leave the active site.

ROLE OF MMPs IN INFLAMMATION, CORONARY REMODELING, AND PLAQUE INSTABILITY

MMPs activity is important for all the phases of an inflammatory response, including initiation, execution and resolution [25]. ECM remodeling by MMPs aids in the initial stages of the inflammatory response by allowing migration of leukocytes into the injured site [54-57]. At the same time, MMPs are also cell-signaling regulators [17] that modify cytokines, chemokines and receptors [58-64] and are able to release antiapoptotic or antiangiogenic factors from the ECM, which may help in the resolution of an inflammatory response [13].

The ECM provides the structural and functional platform of the arterial wall, so it is not surprising that alterations in its turnover, mediated by the activity of MMPs, play a key role throughout the natural history of coronary atherosclerosis, from plaque development and progression to fibrous cap disruption [65, 66]. In particular, MMP-1 (collagenase-1) [65-72], MMP-2 (gelatinase-A) [73, 74], MMP-3 (stromelysin-1) [67, 71, 75, 76], MMP-7 (matrilysin-1) [29], MMP-8 (collagenase-2) [77, 78], MMP-9 (gelatinase-B) [67, 71, 78, 79], MMP-10 (stromelysin-2) [80], MMP-11 [81], MMP-12 (metalloelastase) [29, 72, 82], MMP-13 (collagenase-3) [70, 72], MMP-14 (MT1-MMP) [83, 84], and MMP-16 (MT3-MMP) [85] levels are increased in human atherosclerotic plaques, especially at the macrophage-rich shoulder regions. Interestingly, plaque shoulders and regions of foam cell accumulation contain the highest levels of MMP-9 [67, 86].

As aforementioned, intimal thickening implies the generation of new tissue at least in part by means of hyperplasia and migration of VSMC derived from the media. Removal of the basement membrane and subsequent exposure of the interstitial matrix to the VSMC is promoted by MMPs. This seems to enable a shift from quiescent, contractile VSMC to cells able to migrate and proliferate and eventually mediate repair. The generation of new ECM that favors VSMC migration and proliferation is also promoted by specific MMPs.

Outward (positive) remodeling of coronary vessels has been initially regarded as beneficial by preventing lumen encroachment owing to plaque growth, and hence improving coronary flow [87]. Notwithstanding, several studies have subsequently shown increased levels of inflammatory markers, larger lipid cores and pronounced medial thinning in positive remodeled vessels; being all factors related to the tendency of plaques to undergo rupture [88-91]. Vascular remodeling implies the degradation and reorganization of ECM lead by MMPs [92]. Such phenomenon is already evident at very early stages of atherosclerosis [93] and has a key role in the pathogenesis of plaque disruption [7].

Several clinical studies have established that genetic polymorphism in a variety of MMPs might be useful in determining individual susceptibility to ACS and the extent of coronary atherosclerosis, and are associated with MMPs plasma levels [94-96]. Indeed, Liu *et al.* demonstrated that MMP-3 5A/6A polymorphism was independently associated with the risk of ACS, MMP-3 activity and angiographical severity of coronary atherosclerosis [97, 98].

In parallel, MMP-8 gene variation has also been associated with the extent of coronary atherosclerosis and VCAM-1 levels [99]. Furthermore, Cheng *et al.* recently established that intra-plaque hemorrhage and collagen breakdown in vulnerable atherosclerotic lesions is mediated by activation of MMP-8 and MMP-13 [100].

MMP INHIBITION AS A POTENTIALLY EFFECTIVE ANTIATHEROSCLEROTIC STRATEGY

Specific MMPs have different roles in the development and phenotype of atherosclerosis, and sometimes also contradicting roles, being both beneficial and detrimental [25]. For example, MMP-9 appears as harmful and protective depending on the site and the experimental setup [101]. This paradox complicates the use of broad-spectrum MMPis [102, 103]. MMPs may contribute to the formation and growth of atherosclerotic lesions by facilitating migration of VSMC through the internal elastic lamina into the intima space [104], by favoring monocyte infiltration of the

vascular wall [66], and by triggering fibrous cap rupture. Conversely, MMPs may also aid in the resolution of a plaque by degrading ECM in the intima [104-107].

The expression of MMPs can be induced by a number of inflammatory cytokines, hormones, growth factors and thrombin [76, 85, 108-114]. In particular, C-reactive protein (CRP), an inflammatory marker of atherosclerotic risk, induces the expression of MMP-1 and MMP-10 in macrophages [115] and endothelial cells, contributing to plaque vulnerability [80]. MMP-10 also appears as a useful marker of subclinical atherosclerosis in asymptomatic patients, because its levels are associated with inflammatory markers, increased thickness of the intima media of the carotid artery and presence of atherosclerotic plaques [116]. Furthermore, overexpression of MMP-1 and MMP-9 by macrophages and VSMC has been associated with the pathology and progression of vulnerable lesions [67, 70].

There is abundant evidence that suggests MMP-12 is a harmful protease that favors atherosclerotic plaque development and destabilization. Overexpression of MMP-12 in transgenic rabbits promotes macrophage infiltration and disruption of the internal elastic lamina, accelerating the atherosclerotic process [117]. Animal models have also shown a role of MMP-12 in macrophage recruitment to sites of preinduced inflammation, suggesting MMP-12 as a key player of inflammation [118-121]. Moreover, studies on apolipoprotein E-deficient (apoE(-/-)) and MMP-12 double-knockout mice suggest that MMP-12 may act as a destructive protease that promotes plaque instability by increasing the atherosclerotic lesion size and macrophage content, and decreasing the number of VSMC [122].

On the other hand, there is overwhelming evidence of gelatinase induction and activation in animal models of neointima formation after vascular injury that correlates with the activation and migration of VSMC [10]. These studies have shown upregulation of both MMP-2 and MMP-9 after balloon injury in rat [123, 124], pig [125], baboon [126], rabbit [127], and mouse [128, 129] arteries. Migration and proliferation of VSMC can favor fibrous cap formation and plaque stability [101, 130]. By using apoE/MMP-2 knockout mice it has been demonstrated that MMP-2 contributes to the formation and growth of the fibrous cap in the aortic root [131]. In agreement, increased MMP-2 activity levels were associated with VSMC content and a fibrous phenotype in carotid arteries, suggesting that MMP-2 expression is associated with a stable lesion phenotype [78].

Less clear is the role of MMP-9 in plaque stability based on apoE/MMP double-knockout mice models, suggesting a dual effect [101]. MMP-9 increases lesion size, macrophage content and medial destruction at the base of plaques in the descending aorta, suggesting that MMP-9 promotes instability at this site [132]. However, MMP-9 appears as a protective protease in mouse brachiocephalic arteries, because its presence results in smaller lesions with more smooth muscle content and less macrophage infiltration, promoting plaque stability [122].

Immunopositive MMP-2 and MMP-9 are increased in positive remodeled sections compared to negative remodeled sections [133]. Similarly, increased MMP-2 and MMP-9

levels were found in abdominal aortic aneurysms, an extreme kind of positive remodeling [134, 135]. In parallel, matrix degradation of the fibrous cap shoulder was concomitantly associated with overexpression of MMPs, thereby promoting the vulnerability of atherosclerotic plaques [67].

Ex vivo and *in vivo* studies have shown that both rupture-prone plaques and plaque rupture are highly prevalent even in stable patients [46, 47, 136, 137]. During the past 10 years, the role of MMPs in plaque rupture has become focus of attention since the proteolytic disruption of the fibrous cap overlying a lipid-rich plaque has been established as the most common physiopathological substrate of sudden cardiac death [7, 70, 138].

It has been suggested that one of the main circulating markers of ECM breakdown is MMP-9 [66]. MMP-9 plasma levels are significantly higher in patients with ACS, and correlate with a narrowing of the arterial lumen and restenosis after stent deployment [139, 140]. The serum levels of MMP-9 are significantly higher in patients with CAD with respect to control patients, and correlate directly with those of CRP, interleukin-6 and fibrinogen [141]. To note, MMP-9 expression is increased in plaques of patients with unstable angina with respect to those with stable angina [142, 143]. Furthermore, MMP-9 has been demonstrated to be a predictor of cardiovascular death in patients with coronary heart disease [144], and of ischemic heart disease and high pressure in patients with no history of cardiovascular disease [145]. Altogether, these results place MMP-9 as a possible marker of inflammation in patients with known CAD [141], and suggest a positive relationship between MMP-9 expression and plaque instability and rupture [10]. However, a clinical trial with 389 patients showed that TIMP-1 and not MMP-9 was able to independently predict death and MI [146]. Interestingly, others have suggested that the increase in plasma MMP-9 concentration after an acute coronary event might represent a healing response that involves the recruitment of VSMC rather than the initial of plaque rupture [122].

INHIBITION OF MMPs WITH SMALL MOLECULES

Most synthetic MMPis present some cross-reactivity because they competitively target the structurally-conserved substrate binding pocket [18]. They generally contain a Zn chelating group (hydroxamate, carboxylate, thiolate, phosphinate) and a peptidomimetic moiety that mimics the peptide backbone of the substrate that interacts with the active site [18, 147]. Third generation MMPis have been designed with new Zn-binding groups (pyrone, thiirane) and even without any Zn-coordinating element [148]. The later were designed to target allosteric sites of the MMPs structures by exploiting the recognized flexibility of the MMPs active site [149]. However, they still bind to the S₁' pocket and compete for substrate binding regions [148].

An important and more recent group of MMPis corresponds to tetracyclines and chemically modified tetracyclines (CMTs), which can exhibit antimicrobial or non-antimicrobial activities [25]. Tetracyclines and CMTs can inhibit MMP activity and connective tissue breakdown both *in vitro* and *in vivo* [150]. They have been found to inhibit gelatinases, stromelysins, collagenases and MT-MMPs [150]

from numerous tissue and cellular sources [151]. Because of their chemical nature, these compounds may be able to cross anatomical barriers such as the blood brain barrier and blood retina barrier [25]. CMTs have been extensively studied in a number of animal models of periodontitis [152], metastasis [153], multiple sclerosis [154], and adjuvant arthritis [155].

Nearly 60 MMPs have been developed and tested as clinical candidates over the past 30 years, but, except doxycycline, all of them have failed due to poor safety and a lack of efficacy [147, 156-158]. For example, batimastat (British Biotech), ilomastat (GlycoMed), solimastat (British Biotech) and marimastat (British Biotech) are all broad-spectrum hydroxamate-based peptidomimetic MMPs that were discontinued in Phase I, II or III studies due to severe side effects, including musculoskeletal syndrome [158]. On the other hand, Other MMPs are currently being tested in clinical trials, such as incyclinide (Metastat[®]), S-3304 (Shionogi) and CPA-926 (Kureha Chemical Industry) [147]. Incyclinide is a CMT MMP that inhibits MMP-2 and MMP-9, and is currently in Phase II trials for the treatment of acne, brain tumors, solid tumors, and HIV-related Kaposi's sarcoma, and presents mild to moderate side effects [147, 159]. S-3304 is a novel D-tryptophan derivative MMP that inhibits most potently the activities of MMP-2 and MMP-9 without inhibiting MMP-1, -3 and -7, and has shown promising results in Phase I and II trials for the treatment of lung cancer and solid tumors [147, 160]. Finally, CPA-926 (a pro-drug of Esculetin) is a non-peptidomimetic MMP that is currently in Phase II trials for the treatment of osteoarthritis [161, 162].

Doxycycline: Pharmacokinetics, Pharmacodynamics and Adverse Effects

Doxycycline (Periostat[®]) (Fig. (3)), an antimicrobial CMT, is the only FDA-approved MMP inhibitor used for the treatment of periodontitis [23]. Doxycycline is a reversible, noncompetitive and broad-spectrum MMP inhibitor that binds to allosteric sites proximal to the structural Zn and/or Ca atoms [163, 164]. The proposed mechanism of action results from its ability to chelate these structural ions, which are required to maintain proper enzyme conformation and activity [150, 164]. In the case of MMP-7, it has been reported that doxycycline binds to the enzyme *in vitro* with a stoichiometry of 2.3 ± 0.2 and a dissociation constant of $73 \pm 8 \mu\text{M}$ [164]. Its binding in both pro and active MMPs results in the disruption of the normal conformation of the protein structure [164], leaving the enzymes inactive [150].

Doxycycline inhibitory effect has been tested in both humans and animals in a number of conditions associated with elevated MMP activity, including arthritis [165-168], periodontitis [169, 170] and chronic wounds [171].

Doxycycline is available in oral and intravenous formulations [172], and it is believed to be almost completely absorbed in the duodenum [173], with a bioavailability between 73 and 95% [173, 174], significantly more than other tetracyclines. The half-life absorption ranges from 1 to 2 hours when administered while fasting [175], and the peak concentration varies with dose, being 15.3mg/L ($\sim 30 \mu\text{M}$) after an oral dose of 500mg [176].

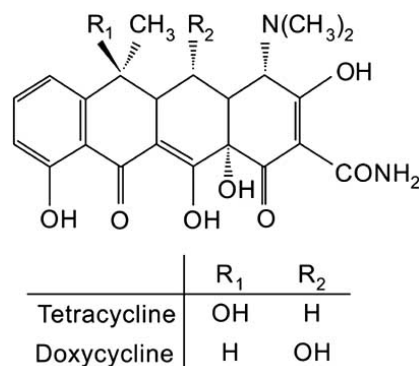


Fig. (3). Chemical structure of tetracycline and doxycycline.

Regarding tissue penetration, doxycycline levels are poor in saliva, below those of serum in bone, skin, fat, tendons and muscle [172, 177], and highest in the liver, kidney and digestive tract [173]. Doxycycline is eliminated unchanged by both the renal and biliary routes [172]; bile concentrations may be 10-25 times greater than serum [178]. About 35-60 % is excreted in urine and the remainder in faeces [179-182]. Doxycycline is slowly absorbed orally. It reaches peak concentration in 2-3 hours, and the elimination half-life ranges between 12 to 25 hours [172]. The area under the serum time curve for a 200mg/day oral dose varies from 41-123mg.h/L and 61-112mg.h/L for intravenous doses [172].

Different studies have been performed to assess the pharmacodynamics of doxycycline as an antimicrobial drug using the minimum inhibitory concentration as a measure of drug potency [172]. With respect to its inhibitory effect on MMPs, it has been shown that doxycycline inhibits collagenase activity more effectively than tetracycline or other CMTs [151]. For example, the concentration of doxycycline required to inhibit 50 % of collagenase activity (IC₅₀) *in vitro* was ~ 13 and ~ 23 times smaller than minocycline and tetracycline, respectively [183]. *In vitro*, the doxycycline IC₅₀ against a colorimetric peptide substrate was reported to be $28 \pm 5 \mu\text{M}$ for MMP-7 [164] and $90 \pm 13 \mu\text{M}$ for MMP-2 [184].

The inhibitory effect of doxycycline varies between different MMPs, as tested *in vitro* by Smith *et al.* In this study, 30 μM doxycycline (value comparable to the concentrations achieved in serum after oral administration) was able to inhibit MMP-1, MMP-8 and MMP-13 activity against type II collagen by 5, 50 and 60% [163].

Mild but relatively common adverse effects of oral doxycycline include hives, shortness of breath, swelling of the face, lip, tongue or throat, headache, dizziness, fever, chills, rash, nausea, vomiting, diarrhea, thrush, vaginitis and photosensitivity [185].

EFFECT OF DOXYCYCLINE ON ATHEROSCLEROSIS AND DIVERSE INFLAMMATORY PROCESSES

A link between chronic infection, the associated inflammatory processes in the periodontal tissue and cardiovascular disease has been undoubtedly established. The presence of carotid artery plaque is associated with

periodontitis and tooth loss [186, 187]. Indeed, using serial carotid ultrasound, Schillinger *et al.* demonstrated that a variety of markers of periodontal disease predict carotid plaque progression, independently of traditional cardiovascular risk factors and the baseline degree of stenosis [188]. Furthermore, a Danish investigation has shown a six-fold increase in the risk of coronary artery disease in individuals with more than 4mm of alveolar bone loss [189]. The beneficial effects of doxycycline in periodontal disease, along with the reported association between periodontitis and atherosclerosis, warrant further research evaluating the effect of doxycycline on atherosclerosis.

There is robust evidence about the detrimental role of positive remodeling in the natural history of coronary atherosclerosis [190]. Several studies have demonstrated an association between coronary remodeling and plaque composition [88, 89, 91]. Indeed, positive remodeling has been established as a major criteria of plaque vulnerability. Since MMPs play a major role in vascular remodeling, MMP inhibition with doxycycline shows promise towards plaque stabilization.

Abdominal aortic aneurysm is an extreme form of vascular remodeling and, as such, nonspecific MMP inhibition has been shown to retard expansive aortic remodeling [102].

Doxycycline effect on vascular remodeling and atherosclerosis has been found to be independent of its antimicrobial properties, as suggested in clinical studies where sub-antimicrobial doses of doxycycline (SDD) decreased the growth rate of abdominal aortic aneurysms [191]. Indeed, Manning *et al.* have shown a significant reduction in abdominal aortic aneurysm formation and severity with doxycycline administration independently from lipid levels obtained or systolic blood pressure [192].

A recent prospective, randomized study including patients requiring carotid endarterectomy, has demonstrated that doxycycline penetrates atherosclerotic plaques at acceptable tissue levels and achieves a significant *in situ* MMP inhibition [193].

The effect of doxycycline on atherosclerosis has been explored by Bendeck *et al.* who demonstrated a significant effect on cell proliferation, migration and MMP activity [194], although it should be noted that a restenosis model (balloon injury in left common carotid of male Sprague-Dawley rats) was used in that study, while such models are not applicable to advanced natural history atherosclerosis, as commonly seen in human. More recently, Madan *et al.* demonstrated a significant decrease in pro-inflammatory cytokines resulting in reduction of atherosclerosis in apoE double knockout mice inoculated with *Porphyromonas gingivalis*, a pathogen related to periodontal disease and systemic inflammation [195].

The MIDAS trial was a prospective, randomized, double-blinded, placebo-controlled pilot study that evaluated the effect SDD in a very small population of ACS patients. At 6 months, although the study was unable to detect an effect on clinical outcome, high-sensitivity CRP was reduced by 46% and pro-MMP-9 activity was reduced by 50% with SDD,

whereas no significant reductions were detected with placebo [196].

In the setting of acute MI and heart failure, upregulation of diverse MMPs has been associated with left ventricular remodeling [197, 198]. Furthermore, it has been ascribed to MMPs a rapid effect on cellular transduction processes before changes in ECM occur, particularly noted on MMP-mediated platelet aggregation [199, 200]. This has led to the exploration and further confirmation that myocardium subjected to ischemia-reperfusion injury releases MMP-2 and that its liberation, of pathological significance for the development of mechanical dysfunction, might be mitigated by doxycycline [201].

CURRENT & FUTURE DEVELOPMENTS

Despite lasting debate about the role of infection in atherosclerosis [202], the largest antibiotic treatment trial to date has failed to show any benefit of azithromycin in post-MI patients with elevated *Chlamydia pneumoniae* titers [203]. In accordance, subsequent randomized trials failed to show benefit of antibiotics in both patients with stable CAD and ACS [204, 205]. These findings are highlighted by the fact that the cardiovascular effects observed with doxycycline were obtained using sub-antimicrobial doses.

Throughout this review, we have outlined the pivotal role of MMPs in the natural history of atherosclerosis and its cardiovascular consequences. Non-selective MMP inhibition with doxycycline appears as a potential strategy to reduce the residual risk observed in patients already at intensive lipid lowering strategies. However, as aforementioned, specific MMPs have different and even contradicting roles in the natural history of atherosclerosis, rendering broad spectrum MMP inhibition an important yet somewhat simplistic approach towards residual risk reduction in coronary atherosclerosis. The effect of a broad-spectrum MMP inhibition is probably expected to be dependent on its degree of inhibition of specific MMPs [103]. It should be stressed though that, overall, the balance of non-selective MMP inhibition might shift to the favorable side only in particular settings such as in ACS, where in addition to its potential plaque stabilization properties, doxycycline shows promise in preventing ischemia-reperfusion injury and left ventricular remodeling. In turn, it seems unlikely that long-term administration of doxycycline might be beneficial in stable angina patients.

Doxycycline appears to be tolerated well by both mice and human, with no significant changes in body weight, lipid metabolism, or blood pressure [192]. From January 1998 to August 2003, approximately 50 million new doxycycline prescriptions were dispensed in the United States, with an event rate of 13 per million [206]. Indeed, long term administration of doxycycline appears to be safer than minocycline, which might be rarely associated with serious adverse events including hypersensitivity syndrome reaction and drug-induced lupus, whereas most common adverse events with doxycycline are mild and gastrointestinal [206, 207].

Nevertheless, to date, most animal models used do not represent advanced coronary atherosclerosis seen in humans,

and clinical studies exploring the role of doxycycline on atherosclerosis progression, as well as powered clinical outcome studies are lacking.

Finally, it is noteworthy that the complexity of MMPs as targets for inhibition, with some MMPs being attributed a potential beneficial effect particularly during the healing response, warrants further research towards the design of selective inhibitors of individual MMPs [208-222].

CONFLICTS OF INTEREST

The authors have no conflict of interest to declare.

ACKNOWLEDGEMENTS

We would like to thank Alejandro Crespo for his assistance and critical review of the manuscript.

ABBREVIATIONS

MMPs	=	Matrix metalloproteinases
CAD	=	Coronary artery disease
MI	=	Myocardial infarction
ACS	=	Acute coronary syndromes
ECM	=	Extracellular matrix
TIMPs	=	Tissue inhibitors of MMPs
VSCM	=	Vascular smooth muscle cells
CRP	=	C-Reactive protein
SDD	=	Sub-antimicrobial doses of doxycycline
CMTs	=	Chemically modified tetracyclines

REFERENCES

- Yusuf S, Reddy S, Ounpuu S, Anand S. Global burden of cardiovascular diseases: Part II: Variations in cardiovascular disease by specific ethnic groups and geographic regions and prevention strategies. *Circulation* 2001; 104(23): 2855-64.
- Little WC, Constantinescu M, Applegate RJ, Kutcher MA, Burrows MT, Kahl FR, *et al.* Can coronary angiography predict the site of a subsequent myocardial infarction in patients with mild-to-moderate coronary artery disease? *Circulation* 1988; 78(5 Pt 1): 1157-66.
- [No authors listed] Randomised trial of cholesterol lowering in 4444 patients with coronary heart disease: The Scandinavian Simvastatin Survival Study (4S). *Lancet* 1994; 344(8934): 1383-9.
- Sacks FM, Pfeffer MA, Moye LA, Rouleau JL, Rutherford JD, Cole TG, *et al.* The effect of pravastatin on coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events Trial investigators. *N Engl J Med* 1996; 335(14):1001-9.
- The Long-Term Intervention with Pravastatin in Ischaemic Disease (LIPID) Study Group. Prevention of cardiovascular events and death with pravastatin in patients with coronary heart disease and a broad range of initial cholesterol levels. *N Engl J Med* 1998; 339(19): 1349-57.
- Kannel WB, Doyle JT, McNamara PM, Quickenon P, Gordon T. Precursors of sudden coronary death. Factors related to the incidence of sudden death. *Circulation* 1975; 51(4): 606-13.
- Falk E, Shah PK, Fuster V. Coronary plaque disruption. *Circulation* 1995; 92(3): 657-71.
- Schwartz GG, Olsson AG, Ezekowitz MD, Ganz P, Oliver MF, Waters D, *et al.* Effects of atorvastatin on early recurrent ischemic events in acute coronary syndromes: The MIRACL study: A randomized controlled trial. *JAMA* 2001; 285(13): 1711-8.
- Bayturan O, Kapadia S, Nicholls SJ, Tuzcu EM, Shao M, Uno K, *et al.* Clinical predictors of plaque progression despite very low levels of low-density lipoprotein cholesterol. *J Am Coll Cardiol* 2010; 55(24): 2736-42.
- Newby AC. Dual role of matrix metalloproteinases (matrixins) in intimal thickening and atherosclerotic plaque rupture. *Physiol Rev* 2005; 85(1): 1-31.
- Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Ann Rev Cell Dev Biol* 2001; 17: 463-516.
- Overall CM, Kleinfeld O. Tumour microenvironment - opinion: Validating matrix metalloproteinases as drug targets and anti-targets for cancer therapy. *Nat Rev Cancer* 2006; 6(3): 227-39.
- Rodriguez D, Morrison CJ, Overall CM. Matrix metalloproteinases: What do they not do? New substrates and biological roles identified by murine models and proteomics. *Biochim Biophys Acta* 2010; 1803(1): 39-54.
- Kessenbrock K, Plaks V, Werb Z. Matrix metalloproteinases: Regulators of the tumor micro environment. *Cell* 2010; 141(1): 52-67.
- Visse R, Nagase H. Matrix metalloproteinases and tissue inhibitors of metalloproteinases: Structure, function, and biochemistry. *Circ Res* 2003; 92(8): 827-39.
- McCawley LJ, Matrisian LM. Matrix metalloproteinases: They're not just for matrix anymore. *Curr Opin Cell Biol* 2001; 13(5): 534-40.
- Overall CM. Dilating the degradome: Matrix metalloproteinase 2 (MMP-2) cuts to the heart of the matter. *Biochem J* 2004; 383(Pt. 3): e5-7.
- Whittaker M, Floyd CD, Brown P, Gearing AJ. Design and therapeutic application of matrix metalloproteinase inhibitors. *Chem Rev* 1999; 99(9): 2735-76.
- Puerta DT, Cohen SM. A bioinorganic perspective on matrix metalloproteinase inhibition. *Curr Top Med Chem* 2004; 4(15): 1551-73.
- Bertini, I, Calderone V, Fragai M, Giachetti A, Loconte M, Luchinat C, *et al.* Exploring the subtleties of drug-receptor interactions: The case of matrix metalloproteinases. *J Am Chem Soc* 2007; 129(9): 2466-75.
- Lauer-Fields J, Brew K, Whitehead JK, Li S, Hammer RP, Gregg B. Triple-helical transition state analogues: A new class of selective matrix metalloproteinase inhibitors. *J Am Chem Soc* 2007; 129(34): 10408-17.
- Golub LM, Lee H-M, Ryan ME, Ryan ME, Payne J, Sorsa T, *et al.* Tetracyclines inhibit connective tissue breakdown by multiple non-antimicrobial mechanisms. *Adv Dent Res* 1998; 12(2): 12-26.
- Peterson JT. Matrix metalloproteinase inhibitor development and the remodeling of drug discovery. *Heart Fail Rev* 2004; 9(1): 63-79.
- Pirard B. Insight into the structural determinants for selective inhibition of matrix metalloproteinases. *Drug Discov Today* 2007; 12(15-16): 640-6.
- Hu J, Van den Steen PE, Sang QA, Opdenakker G. Matrix metalloproteinase inhibitors as therapy for inflammatory and vascular diseases. *Nat Rev Drug Discov* 2007; 6(6): 480-98.
- Overall CM, Lopez-Otin C. Strategies for MMP inhibition in cancer: Innovations for the post-trial era. *Nat Rev Cancer* 2002; 2(9): 657-72.
- Coussens LM, Fingleton B, Matrisian LM, Matrix metalloproteinase inhibitors and cancer: trials and tribulations. *Science* 2002; 295(5564): 2387-92.
- Mackay AR, Gomez DE, Nason AM, Thorgeirsson UP. Studies on the effects of laminin, E-8 fragment of laminin and synthetic laminin peptides PA22-2 and YIGSR on matrix metalloproteinases and tissue inhibitor of metalloproteinase expression. *Lab Invest* 1994; 70(6): 800-6.
- Halpert I, Sires UI, Roby JD, Potter-Perigo S, Wight TN, Shapiro SD, *et al.* Matrilysin is expressed by lipid-laden macrophages at sites of potential rupture in atherosclerotic lesions and localizes to areas of versican deposition, a proteoglycan substrate for the enzyme. *Proc Natl Acad Sci USA* 1996; 93(18): 9748-53.
- Overall CM. Molecular determinants of metalloproteinase substrate specificity: Matrix metalloproteinase substrate binding domains, modules, and exosites. *Mol Biotechnol* 2002; 22(1): 51-86.
- Nagase H, Visse R, Murphy G. Structure and function of matrix metalloproteinases and TIMPs. *Cardiovasc Res* 2006; 69(3): 562-73.

- [32] Maskos K. Crystal structures of MMPs in complex with physiological and pharmacological inhibitors. *Biochimie* 2005; 87(3-4): 249-63.
- [33] Van Wart HE, Birkedal-Hansen H. The cysteine switch: A principle of regulation of metalloproteinase activity with potential applicability to the entire matrix metalloproteinase gene family. *Proc Natl Acad Sci USA* 1990; 87(14): 5578-82.
- [34] Loftus IM, Naylor AR, Bell PR, Thompson MM. Matrix metalloproteinases and atherosclerotic plaque instability. *Br J Surg* 2002; 89(6): 680-94.
- [35] Proudfoot D, Skepper JN, Weissberg PL, Shanahan CM. Acetylated low-density lipoprotein stimulates human vascular smooth muscle cell calcification by promoting osteoblastic differentiation and inhibiting phagocytosis. *Circulation* 2002; 106(24): 3044-50.
- [36] Timpl R. Macromolecular organization of basement membranes. *Curr Opin Cell Biol* 1996; 8(5): 618-24.
- [37] Woods A, Oh ES, Couchman JR. Syndecan proteoglycans and cell adhesion. *Matrix Biol* 1998; 17(7): 477-83.
- [38] Thyberg J, Blomgren K, Roy J, Tran PK, Hedin U. Phenotypic modulation of smooth muscle cells after arterial injury is associated with changes in the distribution of laminin and fibronectin. *J Histochem Cytochem* 1997; 45(6): 837-46.
- [39] Zaleski A, Shi Y, Johnson AG. Diverse origin of intimal cells: Smooth muscle cells, myofibroblasts, fibroblasts, and beyond? *Circ Res* 2002; 91(8): 652-5.
- [40] Newby AC. Vitronectin is implicated as the matrix takes control of neointima formation. *Cardiovasc Res* 2002; 53(4): 779-81.
- [41] Wight TN. The extracellular matrix and atherosclerosis. *Curr Opin Lipidol* 1995; 6(5): 326-34.
- [42] Clowes AW, Clowes MM, Reidy MA. Kinetics of cellular proliferation after arterial injury. III. Endothelial and smooth muscle growth in chronically denuded vessels. *Lab Invest* 1986; 54(3): 295-303.
- [43] Ross R. Atherosclerosis--an inflammatory disease. *N Engl J Med* 1999; 340(2): 115-26.
- [44] Stary HC. The sequence of cell and matrix changes in atherosclerotic lesions of coronary arteries in the first forty years of life. *Eur Heart J* 1990; 11 (Suppl E): 3-19.
- [45] Libby P. Atherosclerosis: Disease biology affecting the coronary vasculature. *Am J Cardiol* 2006; 98(12A): 3Q-9Q.
- [46] Virmani R, Kolodgie FD, Burke AP, Farb A, Schwartz SM. Lessons from sudden coronary death: A comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler Thromb Vasc Biol* 2000; 20(5): 1262-75.
- [47] Arbustini E, Grasso M, Diegoli M, Morbini P, Aguzzi A, Fasanì R, *et al.* Coronary thrombosis in non-cardiac death. *Coron Artery Dis* 1993; 4(9): 751-9.
- [48] Mann J, Davies MJ. Mechanisms of progression in native coronary artery disease: Role of healed plaque disruption. *Heart* 1999; 82(3): 265-8.
- [49] Burke AP, Kolodgie FD, Farb A, Weber DK, Malcom GT, Smialek J, *et al.* Healed plaque ruptures and sudden coronary death: Evidence that subclinical rupture has a role in plaque progression. *Circulation* 2001; 103(7): 934-40.
- [50] Davies MJ, Richardson PD, Woolf N, Katz DR, Mann J. Risk of thrombosis in human atherosclerotic plaques: Role of extracellular lipid, macrophage, and smooth muscle cell content. *Br Heart J* 1993; 69(5): 377-81.
- [51] Gertz SD, Roberts WC. Hemodynamic shear force in rupture of coronary arterial atherosclerotic plaques. *Am J Cardiol* 1990; 66(19): 1368-72.
- [52] Davies MJ. The pathophysiology of acute coronary syndromes. *Heart* 2000; 83(3): 361-6.
- [53] Libby P, Aikawa M. Stabilization of atherosclerotic plaques: New mechanisms and clinical targets. *Nat Med* 2002; 8(11): 1257-62.
- [54] Opdenakker G, Fibbe WE, Van Damme J. The molecular basis of leukocytosis. *Immunol Today* 1998; 19(4): 182-9.
- [55] Puijitt JF, Fibbe WE, Laterveer L, Pieters RA, Lindley IJD, Paemen L, *et al.* Prevention of interleukin-8-induced mobilization of hematopoietic progenitor cells in rhesus monkeys by inhibitory antibodies against the metalloproteinase gelatinase B (MMP-9). *Proc Natl Acad Sci USA* 1999; 96(19): 10863-8.
- [56] Parks WC, Wilson CL, Lopez-Boado YS. Matrix metalloproteinases as modulators of inflammation and innate immunity. *Nat Rev Immunol* 2004; 4(8): 617-29.
- [57] Kollet O, Dar A, Shivtiel S, Kalinkovich A, Lapid K, Sztainberg Y, *et al.* Osteoclasts degrade endosteal components and promote mobilization of hematopoietic progenitor cells. *Nat Med* 2006; 12(6): 657-64.
- [58] Wilson CL, Ouellette AJ, Satchell DP, Ayabe T, López-Boado YS, Stratman JL, *et al.* Regulation of intestinal alpha-defensin activation by the metalloproteinase matrilysin in innate host defense. *Science* 1999; 286(5437): 113-7.
- [59] Van den Steen PE, Proost P, Wuyts A, Van Damme J, Opdenakker G. Neutrophil gelatinase B potentiates interleukin-8 tenfold by aminoterminal processing, whereas it degrades CTAP-III, PF-4, and GRO-alpha and leaves RANTES and MCP-2 intact. *Blood* 2000; 96(8): 2673-81.
- [60] McQuibban GA, Gong JH, Tam EM, McCulloch CA, Clark-Lewis I, Overall CM, *et al.* Inflammation dampened by gelatinase A cleavage of monocyte chemoattractant protein-3. *Science* 2000; 289(5482): 1202-6.
- [61] McQuibban GA, Gong JH, Wong JP, Wallace JL, Clark-Lewis I, Overall CM. Matrix metalloproteinase processing of monocyte chemoattractant proteins generates CC chemokine receptor antagonists with anti-inflammatory properties *in vivo*. *Blood* 2002; 100(4): 1160-7.
- [62] Van Den Steen PE, Wuyts A, Husson SJ, Proost P, Van Damme J, Opdenakker G. Gelatinase B/MMP-9 and neutrophil collagenase/MMP-8 process the chemokines human GCP-2/CXCL6, ENA-78/CXCL5 and mouse GCP-2/LIX and modulate their physiological activities. *Eur J Biochem* 2003; 270(18): 3739-49.
- [63] Nelissen I, Martens E, Van den Steen PE, Proost P, Ronsse I, Opdenakker G, *et al.* Gelatinase B/matrix metalloproteinase-9 cleaves interferon-beta and is a target for immunotherapy. *Brain* 2003; 126(Pt 6): 1371-81.
- [64] McQuibban GA, Butler GS, Gong J-H, Bendall L, Power C, Clark-Lewis I, *et al.* Matrix metalloproteinase activity inactivates the CX chemokine stromal cell-derived factor-1. *J Biol Chem* 2001; 276(47): 43503-8.
- [65] Libby P, Lee RT. Matrix matters. *Circulation* 2000; 102(16): 1874-6.
- [66] Rodriguez JA, Orbe J, Paramo JA. Metalloproteases, vascular remodeling and atherothrombotic syndromes. *Rev Esp Cardiol* 2007; 60(9): 959-67.
- [67] Galis ZS, Sukhova GK, Lark MW, Libby P. Increased expression of matrix metalloproteinases and matrix degrading activity in vulnerable regions of human atherosclerotic plaques. *J Clin Invest* 1994; 94(6): 2493-503.
- [68] Nikkari ST, O'Brien KD, Ferguson M, Hatsukami T, Welgus HG, Alpers CE, *et al.* Interstitial collagenase (MMP-1) expression in human carotid atherosclerosis. *Circulation* 1995; 92(6): 1393-8.
- [69] Nikkari ST, Geary RL, Hatsukami T, Ferguson M, Forough R, Alpers CE, *et al.* Expression of collagen, interstitial collagenase, and tissue inhibitor of metalloproteinases-1 in restenosis after carotid endarterectomy. *Am J Pathol* 1996; 148(3): 777-83.
- [70] Sukhova GK, Schonbeck U, Rabkin E. Evidence for increased collagenolysis by interstitial collagenases-1 and -3 in vulnerable human atherosclerotic plaques. *Circulation* 1999; 99(19): 2503-9.
- [71] Higashikata T, Yamagishi M, Higashi T, Nagata I, Iihara K, Miyamoto S, *et al.* Altered expression balance of matrix metalloproteinases and their inhibitors in human carotid plaque disruption: Results of quantitative tissue analysis using real-time RT-PCR method. *Atherosclerosis* 2006; 185(1): 165-72.
- [72] Yu Y, Koike T, Kitajima S, Liu E, Morimoto M, Shiomi M, *et al.* Temporal and quantitative analysis of expression of metalloproteinases (MMPs) and their endogenous inhibitors in atherosclerotic lesions. *Histol Histopathol* 2008; 23(12): 1503-16.
- [73] Li Z, Li L, Zielke HR, Cheng L, Xiao R, Crow MT, *et al.* Increased expression of 72-kD type IV collagenase (MMP-2) in human aortic atherosclerotic lesions. *Am J Pathol* 1996; 148(1): 121-8.
- [74] Kieffer P, Giummelly P, Schjoth B, Carteaux JP, Villemot JP, Homebeck W, *et al.* Activation of metalloproteinase-2, loss of matrix scleroprotein content and coronary artery calcification. *Atherosclerosis* 2001; 157(1): 251-4.
- [75] Henney AM, Wakeley PR, Davies MJ, Foster K, Hembry R, Murphy G, *et al.* Localization of stromelysin gene expression in atherosclerotic plaques by *in situ* hybridization. *Proc Natl Acad Sci USA* 1991; 88(18): 8154-8.
- [76] Galis ZS, Muszynski M, Sukhova GK, Simon-Morrissey E, Libby P. Enhanced expression of vascular matrix metalloproteinases

- induced *in vitro* by cytokines and in regions of human atherosclerotic lesions. *Ann NY Acad Sci* 1995; 748: 501-7.
- [77] Molloy KJ, Thompson MM, Jones JL, Schwalbe EC, Bell PRF, Naylor AR, *et al.* Unstable carotid plaques exhibit raised matrix metalloproteinase-8 activity. *Circulation* 2004; 110(3): 337-43.
- [78] Sluijter JP, Pulskens WPC, Schoneveld AH, Velema E, Strijder CF, Moll F, *et al.* Matrix metalloproteinase 2 is associated with stable and matrix metalloproteinases 8 and 9 with vulnerable carotid atherosclerotic lesions: A study in human endarterectomy specimen pointing to a role for different extracellular matrix metalloproteinase inducer glycosylation forms. *Stroke* 2006; 37(1): 235-9.
- [79] Brown DL, Hibbs MS, Kearney M, Loushin C, Isner JM. Identification of 92-kD gelatinase in human coronary atherosclerotic lesions. Association of active enzyme synthesis with unstable angina. *Circulation* 1995; 91(8): 2125-31.
- [80] Montero I, Orbe J, Varo N, Belouqui O, Monreal JI, Rodríguez JA, *et al.* C-reactive protein induces matrix metalloproteinase-1 and -10 in human endothelial cells: Implications for clinical and subclinical atherosclerosis. *J Am Coll Cardiol* 2006; 47(7): 1369-78.
- [81] Schonbeck U, Mach F, Sukhova GK, Atkinson E, Levesque E, Herman M, *et al.* Expression of stromelysin-3 in atherosclerotic lesions: regulation via CD40-CD40 ligand signaling *in vitro* and *in vivo*. *J Exp Med* 1999; 189(5): 843-53.
- [82] Morgan AR, Rerkasem K, Gallagher PJ, Zhang B, Morris GE, Calder PC, *et al.* Differences in matrix metalloproteinase-1 and matrix metalloproteinase-12 transcript levels among carotid atherosclerotic plaques with different histopathological characteristics. *Stroke* 2004; 35(6): 1310-5.
- [83] Rajavashisth TB, Xu X-P, Jovinge S, Meisel S, Xu X-O, Chai N-N, *et al.* Membrane type 1 matrix metalloproteinase expression in human atherosclerotic plaques: Evidence for activation by proinflammatory mediators. *Circulation* 1999; 99(24): 3103-9.
- [84] Johnson JL, Sala-Newby GB, Ismail Y, Aguilera CM, Newby AC. Low tissue inhibitor of metalloproteinases 3 and high matrix metalloproteinase 14 levels defines a subpopulation of highly invasive foam-cell macrophages. *Arterioscler Thromb Vasc Biol* 2008.
- [85] Uzui H, Harpf A, Liu M, Doherty TM, Shukla A, Chai NN, *et al.* Increased expression of membrane type 3-matrix metalloproteinase in human atherosclerotic plaque: Role of activated macrophages and inflammatory cytokines. *Circulation* 2002; 106(24): 3024-30.
- [86] Herman MP, Sukhova GK, Kiesel W, Foster D, Kehry MR, Libby P, *et al.* Tissue factor pathway inhibitor-2 is a novel inhibitor of matrix metalloproteinases with implications for atherosclerosis. *J Clin Invest* 2001; 107(9): 1117-26.
- [87] Glagov S, Weisenberg E, Zarins CK, Stankunavicius R, Kolletis GJ. Compensatory enlargement of human atherosclerotic coronary arteries. *N Engl J Med* 1987; 316(22): 1371-5.
- [88] Pasterkamp G, Schoneveld AH, van der Wal AC, Haudenschild CC, Clarijs RJG, Becker AE, *et al.* Relation of arterial geometry to luminal narrowing and histologic markers for plaque vulnerability: The remodeling paradox. *J Am Coll Cardiol* 1998; 32(3): 655-62.
- [89] Varnava AM, Mills PG, Davies MJ. Relationship between coronary artery remodeling and plaque vulnerability. *Circulation* 2002; 105(8): 939-43.
- [90] Burke AP, Kolodgie FD, Farb A, Weber D, Virmani R. Morphological predictors of arterial remodeling in coronary atherosclerosis. *Circulation* 2002; 105(3): 297-303.
- [91] Rodriguez-Granillo GA, Serruys PW, Garcia-Garcia HM, Aoki J, Valgimigli M, van Mieghem CAG, *et al.* Coronary artery remodeling is related to plaque composition. *Heart* 2006; 92(3): 388-91.
- [92] Galis ZS, Khatri JJ. Matrix metalloproteinases in vascular remodeling and atherogenesis: The good, the bad, and the ugly. *Circ Res* 2002; 90(3): 251-62.
- [93] Rodriguez-Granillo GA, de Winter S, Bruining N, Ligthart JMR, García-García HM, Valgimigli M, *et al.* Effect of perindopril on coronary remodeling: Insights from a multicentre, randomized study. *Eur Heart J* 2007; 28(19): 2326-31.
- [94] Terashima M, Akita H, Kanazawa K, Inoue N, Yamada S, Ito K, *et al.* Stromelysin promoter 5A/6A polymorphism is associated with acute myocardial infarction. *Circulation* 1999; 99(21): 2717-9.
- [95] Ghaderian SM, Najar AR, Panah TAS. Genetic polymorphisms and plasma levels of matrix metalloproteinases and their relationships with developing acute myocardial infarction. *Coronary Artery Dis* 2010; 21(6): 330-5.
- [96] Roman-García P, Coto E, Reguero JR, Cannata-Andía JB, Lozano I, Avanzas P, *et al.* Matrix metalloproteinase 1 promoter polymorphisms and risk of myocardial infarction: A case-control study in a Spanish population. *Coronary Artery Dis* 2009; 20(6): 383-6.
- [97] Liu PY, Li YH, Chan SH, Lin LJ, Wu HL, Shi GY, *et al.* Genotype-phenotype association of matrix metalloproteinase-3 polymorphism and its synergistic effect with smoking on the occurrence of acute coronary syndrome. *Am J Cardiol* 2006; 98(8): 1012-7.
- [98] Abilleira S, Bevan S, Markus HS. The role of genetic variants of matrix metalloproteinases in coronary and carotid atherosclerosis. *J Med Genet* 2006; 43(12): 897-901.
- [99] Laxton RC, Hu Y, Duchene J, Zhang F, Zhang Z, Leung KY, *et al.* A role of matrix metalloproteinase-8 in atherosclerosis. *Circulation Res* 2009; 105(9): 921-9.
- [100] Cheng C, Tempel D, van Haperen R, van Damme L, Algur M, Krams R, *et al.* Activation of MMP8 and MMP13 by angiotensin II correlates to severe intra-plaque hemorrhages and collagen breakdown in atherosclerotic lesions with a vulnerable phenotype. *Atherosclerosis* 2009; 204(1): 26-33.
- [101] Newby AC. Do metalloproteinases destabilize vulnerable atherosclerotic plaques? *Curr Opin Lipidol* 2006; 17(5): 556-61.
- [102] Prescott MF, Sawyer WK, Von Linden-Reed J, Jeune M, Chou M, Caplan SL, *et al.* Effect of matrix metalloproteinase inhibition on progression of atherosclerosis and aneurysm in LDL receptor-deficient mice overexpressing MMP-3, MMP-12, and MMP-13 and on restenosis in rats after balloon injury. *Ann NY Acad Sci* 1999; 878: 179-90.
- [103] Johnson JL, Fritsche-Danielson R, Behrendt M, Westin-Eriksson A, Wennbo H, Herslof M, *et al.* Effect of broad-spectrum matrix metalloproteinase inhibition on atherosclerotic plaque stability. *Cardiovasc Res* 2006; 71(3): 586-95.
- [104] Jones CB, Sane DC, Herrington DM. Matrix metalloproteinases: A review of their structure and role in acute coronary syndrome. *Cardiovasc Res* 2003; 59(4): 812-23.
- [105] Ardans JA, Blum A, Mangan PR, Wientroub S, Cannon RO, Wahl LM, *et al.* Raloxifene-mediated increase in matrix metalloproteinase-1 production by activated monocytes. *Arterioscler Thromb Vasc Biol* 2001; 21(8): 1265-8.
- [106] Silence J, Collen D, Lijnen HR. Reduced atherosclerotic plaque but enhanced aneurysm formation in mice with inactivation of the tissue inhibitor of metalloproteinase-1 (TIMP-1) gene. *Circ Res* 2002; 90(8): 897-903.
- [107] Silence J, Lupu F, Collen D, Lijnen HR. Persistence of atherosclerotic plaque but reduced aneurysm formation in mice with stromelysin-1 (MMP-3) gene inactivation. *Arterioscler Thromb Vasc Biol* 2001; 21(9): 1440-5.
- [108] Galis ZS, Muszynski M, Sukhova GK, Simon-Morrissey E, Unemori EN, Lark MW, *et al.* Cytokine-stimulated human vascular smooth muscle cells synthesize a complement of enzymes required for extracellular matrix digestion. *Circ Res* 1994; 75(1): 181-9.
- [109] Galis ZS, Kranzhöfer R, Fenton JW, Libby P. Thrombin promotes activation of matrix metalloproteinase-2 produced by cultured vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 1997; 17(3): 483-9.
- [110] Duhamel-Clerin E, Orvain C, Lanza F, Cazenave JP, Klein-Soyer C. Thrombin receptor-mediated increase of two matrix metalloproteinases, MMP-1 and MMP-3, in human endothelial cells. *Arterioscler Thromb Vasc Biol* 1997; 17(10): 1931-8.
- [111] Jormsjo S, Ye S, Moritz J, Walter DH, Dimmeler S, Zeiher AM, *et al.* Allele-specific regulation of matrix metalloproteinase-12 gene activity is associated with coronary artery luminal dimensions in diabetic patients with manifest coronary artery disease. *Circ Res* 2000; 86(9): 998-1003.
- [112] Siwik DA, Chang DL, Colucci WS. Interleukin-1beta and tumor necrosis factor-alpha decrease collagen synthesis and increase matrix metalloproteinase activity in cardiac fibroblasts *in vitro*. *Circ Res* 2000; 86(12): 1259-65.
- [113] Feinberg MW, Jain MK, Werner F, Sibinga NE, Wiesel P, Wang H, *et al.* Transforming growth factor-beta 1 inhibits cytokine-mediated induction of human metalloelastase in macrophages. *J Biol Chem* 2000; 275(33): 25766-73.
- [114] Arenas IA, Xu Y, Lopez-Jaramillo P, Davidge ST. Angiotensin II-induced MMP-2 release from endothelial cells is mediated by TNF-alpha. *Am J Physiol Cell Physiol* 2004; 286(4): C779-84.

- [115] Williams TN, Zhang CX, Game BA, He L, Huang Y. C-Reactive protein stimulates MMP-1 expression in U937 histiocytes through Fe[gamma]RII and extracellular signal-regulated kinase pathway: An implication of CRP involvement in plaque destabilization. *Arterioscler Thromb Vasc Biol* 2004; 24(1): 61-6.
- [116] Orbe J, Montero I, Rodríguez JA, Beloqui O, Roncal C, Páramo JA, *et al.* Independent association of matrix metalloproteinase-10, cardiovascular risk factors and subclinical atherosclerosis. *J Thromb Haemost* 2007; 5(1): 91-7.
- [117] Liang J, Liu E, Yu Y, Kitajima S, Koike T, Jin Y, *et al.* Macrophage metalloelastase accelerates the progression of atherosclerosis in transgenic rabbits. *Circulation* 2006; 113(16): 1993-2001.
- [118] Shipley JM, Wesselschmidt RL, Kobayashi DK, Ley TJ, Shapiro SD. Metalloelastase is required for macrophage-mediated proteolysis and matrix invasion in mice. *Proc Natl Acad Sci USA* 1996; 93(9): 3942-6.
- [119] Wang X, Liang J, Koike T, Sun H, Ichikawa T, Kitajima S, *et al.* Overexpression of human matrix metalloproteinase-12 enhances the development of inflammatory arthritis in transgenic rabbits. *Am J Pathol* 2004; 165(4): 1375-83.
- [120] Fan J, Wang X, Wu L, Matsumoto SI, Liang J, Koike T, *et al.* Macrophage-specific overexpression of human matrix metalloproteinase-12 in transgenic rabbits. *Transgenic Res* 2004; 13(3): 261-9.
- [121] Warner RL, Lukacs NW, Shapiro SD, Bhagarvathula N, Nerusu KC, Varani J, *et al.* Role of metalloelastase in a model of allergic lung responses induced by cockroach allergen. *Am J Pathol* 2004; 165(6): 1921-30.
- [122] Johnson JL, George SJ, Newby AC, Jackson CL. Divergent effects of matrix metalloproteinases 3, 7, 9, and 12 on atherosclerotic plaque stability in mouse brachiocephalic arteries. *Proc Natl Acad Sci USA* 2005; 102(43): 15575-80.
- [123] Bendeck MP, Zempo N, Clowes AW, Galaray RE, Reidy MA. Smooth muscle cell migration and matrix metalloproteinase expression after arterial injury in the rat. *Circ Res* 1994; 75(3): 539-45.
- [124] Zempo N, Kenagy RD, Au YP, Bendeck M, Clowes MM, Reidy MA, *et al.* Matrix metalloproteinases of vascular wall cells are increased in balloon-injured rat carotid artery. *J Vasc Surg* 1994; 20(2): 209-17.
- [125] Southgate KM, Fisher M, Banning AP, Thurston VJ, Baker AH, Fabunmi RP, *et al.* Upregulation of basement membrane-degrading metalloproteinase secretion after balloon injury of pig carotid arteries. *Circ Res* 1996; 79(6): 1177-87.
- [126] Kenagy RD, Vergel S, Mattsson E, Bendeck M, Reidy MA, Clowes AW, *et al.* The role of plasminogen, plasminogen activators, and matrix metalloproteinases in primate arterial smooth muscle cell migration. *Arterioscler Thromb Vasc Biol* 1996; 16(11): 1373-82.
- [127] Aoyagi M, Yamamoto M, Azuma H, Nagashima G, Niimi Y, Tamaki M, *et al.* Immunolocalization of matrix metalloproteinases in rabbit carotid arteries after balloon denudation. *Histochem Cell Biol* 1998; 109(2): 97-102.
- [128] Lijnen HR, Lupu F, Moons L, Carmeliet P, Gouling D, Collen D, *et al.* Temporal and topographic matrix metalloproteinase expression after vascular injury in mice. *Thromb Haemost* 1999; 81(5): 799-807.
- [129] Godin D, Ivan E, Johnson C, Magid R, Galis ZS. Remodeling of carotid artery is associated with increased expression of matrix metalloproteinases in mouse blood flow cessation model. *Circulation* 2000; 102(23): 2861-6.
- [130] Newby AC. Matrix metalloproteinases regulate migration, proliferation, and death of vascular smooth muscle cells by degrading matrix and non-matrix substrates. *Cardiovasc Res* 2006; 69(3): 614-24.
- [131] Kuzuya M, Nakamura K, Sasaki T, Cheng XW, Itohar S, Iguchi A, *et al.* Effect of MMP-2 deficiency on atherosclerotic lesion formation in apoE-deficient mice. *Arterioscler Thromb Vasc Biol* 2006; 26(5): 1120-5.
- [132] Lutun A, Lutgens E, Manderveld A, Maris K, Collen D, Carmeliet P, *et al.* Loss of matrix metalloproteinase-9 or matrix metalloproteinase-12 protects apolipoprotein E-deficient mice against atherosclerotic media destruction but differentially affects plaque growth. *Circulation* 2004; 109(11): 1408-14.
- [133] Pasterkamp G, Schoneveld AH, Hijnen DJ, de Kleijn DP, Teepen H, van der Wal AC, *et al.* Atherosclerotic arterial remodeling and the localization of macrophages and matrix metalloproteinases 1, 2 and 9 in the human coronary artery. *Atherosclerosis* 2000; 150(2): 245-53.
- [134] Thompson RW, Parks WC. Role of matrix metalloproteinases in abdominal aortic aneurysms. *Ann NY Acad Sci* 1996; 800: 157-74.
- [135] Knox JB, Sukhova GK, Whittemore AD, Libby P. Evidence for altered balance between matrix metalloproteinases and their inhibitors in human aortic diseases. *Circulation* 1997; 95(1): 205-12.
- [136] Rodriguez-Granillo GA, García-García HM, Valgimigli M, Vaina S, van Mieghem C, van Geuns RJ, *et al.* Global characterization of coronary plaque rupture phenotype using three-vessel intravascular ultrasound radiofrequency data analysis. *Eur Heart J* 2006; 27(16): 1921-7.
- [137] Rodriguez-Granillo GA, García-García HM, Mc Fadden EP, Valgimigli M, Aoki J, de Feyter P, *et al.* *In vivo* intravascular ultrasound-derived thin-cap fibroatheroma detection using ultrasound radiofrequency data analysis. *J Am Coll Cardiol* 2005; 46(11): 2038-42.
- [138] Shah PK, Galis ZS. Matrix metalloproteinase hypothesis of plaque rupture: Players keep piling up but questions remain. *Circulation* 2001; 104(16): 1878-80.
- [139] Kai H, Ikeda H, Yasukawa H, Kai M, Seki Y, Kuwahara F, *et al.* Peripheral blood levels of matrix metalloproteinases-2 and -9 are elevated in patients with acute coronary syndromes. *J Am Coll Cardiol* 1998; 32(2): 368-72.
- [140] Inokubo Y, Hanada H, Ishizaka H, Fukushi T, Kamada T, Okumura K, *et al.* Plasma levels of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 are increased in the coronary circulation in patients with acute coronary syndrome. *Am Heart J* 2001; 141(2): 211-7.
- [141] Ferroni P, Basili S, Martini F, Cardarelli CM, Ceci F, Di Franco M, Bertazzoni G, *et al.* Serum metalloproteinase 9 levels in patients with coronary artery disease: A novel marker of inflammation. *J Investig Med* 2003; 51(5): 295-300.
- [142] Loftus IM, Goodall S, Crowther M, Jones L, Bell PRF, Naylor AR, *et al.* Increased MMP-9 activity in acute carotid plaques: Therapeutic avenues to prevent stroke. *Ann NY Acad Sci* 1999; 878: 551-4.
- [143] Loftus IM, Naylor AR, Goodall S, Crowther M, Jones L, Bell PR, *et al.* Increased matrix metalloproteinase-9 activity in unstable carotid plaques. A potential role in acute plaque disruption. *Stroke* 2000; 31(1): 40-7.
- [144] Blankenberg S, Rupprecht HJ, Poirier O, Bickel C, Smieja M, Hafner G, *et al.* Plasma concentrations and genetic variation of matrix metalloproteinase 9 and prognosis of patients with cardiovascular disease. *Circulation* 2003; 107(12): 1579-85.
- [145] Timms PM, Mannan N, Hitman GA, Noonan K, Mills PG, Syndercombe-court D, *et al.* Circulating MMP9, vitamin D and variation in the TIMP-1 response with VDR genotype: Mechanisms for inflammatory damage in chronic disorders? *QJM* 2002; 95(12): 787-96.
- [146] Cavusoglu E, Ruwende C, Chopra V, Yanamadala S, Eng C, Clark LT, *et al.* Tissue inhibitor of metalloproteinase-1 (TIMP-1) is an independent predictor of all-cause mortality, cardiac mortality, and myocardial infarction. *Am Heart J* 2006; 151(5): 1101. e1-8.
- [147] Dorman G, Cseh S, Hajdu I, Barna L, Konya D, Kupai K, *et al.* Matrix metalloproteinase inhibitors: A critical appraisal of design principles and proposed therapeutic utility. *Drugs* 2010; 70(8): 949-64.
- [148] Overall CM, Kleinfeld O. Towards third generation matrix metalloproteinase inhibitors for cancer therapy. *Br J Cancer* 2006; 94(7): 941-6.
- [149] Bertini I, Calderone V, Cosenza M, Fragai M, Lee Y-M, Luchinat C, *et al.* Conformational variability of matrix metalloproteinases: Beyond a single 3D structure. *Proc Natl Acad Sci USA* 2005; 102(15): 5334-9.
- [150] Acharya MR, Figg VJ, Sparreboom A. Chemically modified tetracyclines as inhibitors of matrix metalloproteinases. *Drug Resist Updat* 2004; 7(3): 195-208.
- [151] Golub LM, Ramamurthy NS, McNamara TF, Greenwald RA, Rifkin BR. Tetracyclines inhibit connective tissue breakdown: New therapeutic implications for an old family of drugs. *Crit Rev Oral Biol Med* 1991; 2(3): 297-321.

- [152] Ramamurthy NS, Rifkin BR, Greenwald RA, Xu JW, Liu Y, Turner G, *et al.* Inhibition of matrix metalloproteinase-mediated periodontal bone loss in rats: A comparison of 6 chemically modified tetracyclines. *J Periodontol* 2002; 73(7): 726-34.
- [153] Lokeshwar BL, Selzer MG, Zhu BQ, Block NL, Golub LM. Inhibition of cell proliferation, invasion, tumor growth and metastasis by an oral non-antimicrobial tetracycline analog (COL-3) in a metastatic prostate cancer model. *Int J Cancer* 2002; 98(2): 297-309.
- [154] Brundula V, Rewcastle NB, Metz LM, Bernard CC, Yong WV. Targeting leukocyte MMPs and transmigration: Minocycline as a potential therapy for multiple sclerosis. *Brain* 2002; 125(Pt 6): 1297-308.
- [155] Zernicke RF, Wohl GR, Greenwald RA, Moak SA, Leng W, Golub LM, *et al.* Administration of systemic matrix metalloproteinase inhibitors maintains bone mechanical integrity in adjuvant arthritis. *J Rheumatol* 1997; 24(7): 1324-31.
- [156] Turk B. Targeting proteases: Successes, failures and future prospects. *Nat Rev Drug Discov* 2006; 5(9): 785-99.
- [157] Fingleton B. Matrix metalloproteinases as valid clinical targets. *Curr Pharm Des* 2007; 13(3): 333-46.
- [158] Fingleton B. MMPs as therapeutic targets--still a viable option? *Semin Cell Dev Biol* 2008; 19(1): 61-8.
- [159] Viera MH, Perez OA, Berman B. Incyclinide. *Drugs Future* 2007; 32(3): 209-14.
- [160] Chiappori AA, Eckhardt SG, Bukowski R, Sullivan DM, Ikeda M, Yano Y, *et al.* A phase I pharmacokinetic and pharmacodynamic study of s-3304, a novel matrix metalloproteinase inhibitor, in patients with advanced and refractory solid tumors. *Clin Cancer Res* 2007; 13(7): 2091-9.
- [161] Yamada H, Watanabe K, Saito T, Hayashi H, Niitani Y, Kikuchi T, *et al.* Esculetin (dihydroxycoumarin) inhibits the production of matrix metalloproteinases in cartilage explants, and oral administration of its prodrug, CPA-926, suppresses cartilage destruction in rabbit experimental osteoarthritis. *J Rheumatol* 1999; 26(3): 654-62.
- [162] Burrage PS, Mix KS, Brinckerhoff CE. Matrix metalloproteinases: Role in arthritis. *Front Biosci* 2006; 11: 529-43.
- [163] Smith GN, Jr., Mickler EA, Hasty KA, Brandt KD. Specificity of inhibition of matrix metalloproteinase activity by doxycycline: Relationship to structure of the enzyme. *Arthritis Rheumatism* 1999; 42(6): 1140-6.
- [164] Garcia RA, Pantazatos DP, Gessner CR, Go KV, Woods VL, Villarreal FJ, *et al.* Molecular interactions between matrilysin and the matrix metalloproteinase inhibitor doxycycline investigated by deuterium exchange mass spectrometry. *Mol Pharmacol* 2005; 67(4): 1128-36.
- [165] Yu LP, Smith GN, Hasty KA, Brandt KD. Doxycycline inhibits type XI collagenolytic activity of extracts from human osteoarthritic cartilage and of gelatinase. *J Rheumatol* 1991; 18(10): 1450-2.
- [166] Lauhio A, Kontinen YT, Salo T, Tschesche H, Nordström D, Lähdevirta J, *et al.* The *in vivo* effect of doxycycline treatment on matrix metalloproteinases in reactive arthritis. *Ann NY Acad Sci* 1994; 732: 431-2.
- [167] Lindy O, Kontinen YT, Sorsa T, Ding Y, Santavirta S, Ceponis A, *et al.* Matrix metalloproteinase 13 (collagenase 3) in human rheumatoid synovium. *Arthritis Rheum* 1997; 40(8): 1391-9.
- [168] Shlopov BV, Smith GN, Cole AA, Hasty KA. Differential patterns of response to doxycycline and transforming growth factor beta1 in the down-regulation of collagenases in osteoarthritic and normal human chondrocytes. *Arthritis Rheum* 1999; 42(4): 719-27.
- [169] Golub LM, Sorsa T, Lee HM, Ciancio S, Sorbi D, Ramamurthy NS, *et al.* Doxycycline inhibits neutrophil (PMN)-type matrix metalloproteinases in human adult periodontitis gingiva. *J Clin Periodontol* 1995; 22(2): 100-9.
- [170] Golub LM, McNamara TF, Ryan ME, Kohut B, Blieden T, Payonk G, *et al.* Adjunctive treatment with subantimicrobial doses of doxycycline: Effects on gingival fluid collagenase activity and attachment loss in adult periodontitis. *J Clin Periodontol* 2001; 28(2): 146-56.
- [171] Siemonsma MA, de Hingh IH, de Man BM, Lomme RM, Verhofstad AA, Hendriks T, *et al.* Doxycycline improves wound strength after intestinal anastomosis in the rat. *Surgery* 2003; 133(3): 268-76.
- [172] Agwuh KN, MacGowan A. Pharmacokinetics and pharmacodynamics of the tetracyclines including glycylicyclines. *J Antimicrob Chemother* 2006; 58(2): 256-65.
- [173] Saivin S, Houin G. Clinical pharmacokinetics of doxycycline and minocycline. *Clin Pharmacokinet* 1988; 15(6): 355-66.
- [174] Dimmling T, Vanderbeke O. Pharmacokinetics after oral and intravenous administration of tetracycline compounds (author's transl). *Med Klin* 1975; 70(7): 279-85.
- [175] Welling PG, Koch PA, Lau CC, Craig WA. Bioavailability of tetracycline and doxycycline in fasted and nonfasted subjects. *Antimicrob Agents Chemother* 1977; 11(3): 462-9.
- [176] Adadevoh BK, Ogunnaike IA, Bolodeoku JO. Serum levels of doxycycline in normal subjects after a single oral dose. *Br Med J* 1976; 1(6014): 880.
- [177] Gnärpe H, Dornbusch K, Hagg O. Doxycycline concentration levels in bone, soft tissue and serum after intravenous infusion of doxycycline. A clinical study. *Scand J Infect Dis* 1976; (Suppl 9): 54-7.
- [178] Alestig K. Studies on doxycycline during intravenous and oral treatment with reference to renal function. *Scand J Infect Dis* 1973; 5(3): 193-8.
- [179] Steigbigel NH, Reed CW, Finland M. Absorption and excretion of five tetracycline analogues in normal young men. *Am J Med Sci* 1968; 255: 296-312.
- [180] Ylitalo P, Hinkka H, Neuvonen PJ. Effect of exercise on the serum level and urinary excretion of tetracycline, doxycycline and sulphamethizole. *Eur J Clin Pharmacol* 1977; 12(5): 367-73.
- [181] Heaney D, Eknoyan G. Minocycline and doxycycline kinetics in chronic renal failure. *Clin Pharmacol Ther* 1978; 24(2): 233-9.
- [182] Campistron G, Coulais Y, Caillard C, Mosser J, Pontagnier H, Houin G, *et al.* Pharmacokinetics and bioavailability of doxycycline in humans. *Arzneimittelforschung* 1986; 36(11): 1705-7.
- [183] Burns FR, Stack MS, Gray RD, Paterson CA. Inhibition of purified collagenase from alkali-burned rabbit corneas. *Invest Ophthalmol Vis Sci* 1989; 30(7): 1569-75.
- [184] Nicolescu AC, Holt A, Kandasamy AD, Pacher P, Schulz R. Inhibition of matrix metalloproteinase-2 by PARP inhibitors. *Biochem Biophys Res Commun* 2009; 387(4): 646-50.
- [185] Stechmiller J, Cowan L, Schultz G. The role of doxycycline as a matrix metalloproteinase inhibitor for the treatment of chronic wounds. *Biol Res Nurs* 11(4): 336-44.
- [186] Beck JD, Elter JR, Heiss G, Couper D, Mauriello SM, Offenbacher S, *et al.* Relationship of periodontal disease to carotid artery intima-media wall thickness: The atherosclerosis risk in communities (ARIC) study. *Arterioscler Thromb Vasc Biol* 2001; 21(11): 1816-22.
- [187] Desvarieux M, Demmer RT, Rundek T, Boden-Albala B, Jacobs DR, Papapanou PN, *et al.* Relationship between periodontal disease, tooth loss, and carotid artery plaque: the oral infections and vascular disease epidemiology study (INVEST). *Stroke* 2003; 34(9): 2120-5.
- [188] Schillinger T, Kluger W, Exner M, Mlekusch W, Sabeti S, Amighi J, *et al.* Dental and periodontal status and risk for progression of carotid atherosclerosis: The inflammation and carotid artery risk for atherosclerosis study dental substudy. *Stroke* 2006; 37(9): 2271-6.
- [189] Geismar K, Stoltze K, Sigurd B, Gyntelberg F, Holmstrup P. Periodontal disease and coronary heart disease. *J Periodontol* 2006; 77(9): 1547-54.
- [190] Schaar JA, Muller JE, Falk E, Virmani R, Fuster V, Serruys PW, *et al.* Terminology for high-risk and vulnerable coronary artery plaques. Report of a meeting on the vulnerable plaque, June 17 and 18, 2003, Santorini, Greece. *Eur Heart J* 2004; 25(12): 1077-82.
- [191] Mosorin M, Juvonen J, Biancari F, Satta J, Surcel HM, Leinonen M, *et al.* Use of doxycycline to decrease the growth rate of abdominal aortic aneurysms: A randomized, double-blind, placebo-controlled pilot study. *J Vasc Surg* 2001; 34(4): 606-10.
- [192] Manning MW, Cassis LA, Daugherty A. Differential effects of doxycycline, a broad-spectrum matrix metalloproteinase inhibitor, on angiotensin II-induced atherosclerosis and abdominal aortic aneurysms. *Arterioscler Thromb Vasc Biol* 2003; 23(3): 483-8.
- [193] Axisa B, Loftus IM, Naylor AR, Goodall S, Jones L, Bell PR, *et al.* Prospective, randomized, double-blind trial investigating the effect of doxycycline on matrix metalloproteinase expression within atherosclerotic carotid plaques. *Stroke* 2002; 33(12): 2858-64.
- [194] Bendeck MP, Conte M, Zhang M, Nili N, Strauss BH, Farwell SM. Doxycycline modulates smooth muscle cell growth, migration, and

- matrix remodeling after arterial injury. *Am J Pathol* 2002; 160(3): 1089-95.
- [195] Madan M, Bishayi B, Hoge M, Messas E, Amar S. Doxycycline affects diet- and bacteria-associated atherosclerosis in an ApoE heterozygote murine model: Cytokine profiling implications. *Atherosclerosis* 2007;190(1): 62-72.
- [196] Brown DL, Desai KK, Vakili BA, Nouneh C, Lee HM, Golub LM. Clinical and biochemical results of the metalloproteinase inhibition with subantimicrobial doses of doxycycline to prevent acute coronary syndromes (MIDAS) pilot trial. *Arterioscler Thromb Vasc Biol* 2004; 24(4): 733-8.
- [197] Spinale FG, Coker ML, Thomas CV, Walker JD, Mukherjee R, Hebbar L. Time-dependent changes in matrix metalloproteinase activity and expression during the progression of congestive heart failure: Relation to ventricular and myocyte function. *Circ Res* 1998; 82(4): 482-95.
- [198] Tyagi SC, Campbell SE, Reddy HK, Tjahja E, Voelker DJ. Matrix metalloproteinase activity expression in infarcted, noninfarcted and dilated cardiomyopathic human hearts. *Mol Cell Biochem* 1996; 155(1):13-21.
- [199] Cleutjens JP. The role of matrix metalloproteinases in heart disease. *Cardiovasc Res* 1996; 32(5): 816-21.
- [200] Sawicki G, Salas E, Murat J, Miszta-Lane H, Radomski MW. Release of gelatinase A during platelet activation mediates aggregation. *Nature* 1997; 386(6625): 616-9.
- [201] Cheung PY, Sawicki G, Wozniak M, Wang W, Radomski MW, Schulz R. Matrix metalloproteinase-2 contributes to ischemia-reperfusion injury in the heart. *Circulation* 2000; 101(15): 1833-9.
- [202] Matturri L, Cazzullo A, Turconi P, Roncoroni L, Grana D, Milei J. Inflammatory cells, apoptosis and *Chlamydia pneumoniae* infection in atherosclerosis. *Int J Cardiol* 2000; 75(1): 23-33; discussion 33-25.
- [203] O'Connor CM, Dunne MW, Pfeffer MA, Muhlestein JB, Yao L, Gupta S, *et al.* Azithromycin for the secondary prevention of coronary heart disease events: The WIZARD study: A randomized controlled trial. *JAMA* 2003; 290(11): 1459-66.
- [204] Grayston JT, Kronmal RA, Jackson LA, Parisi AF, Muhlestein JB, Cohen JD, *et al.* Azithromycin for the secondary prevention of coronary events. *N Engl J Med* 2005; 352(16): 1637-45.
- [205] Cannon CP, Braunwald E, McCabe CH, Grayston JT, Muhlestein B, Giugliano RP, *et al.* Antibiotic treatment of *Chlamydia pneumoniae* after acute coronary syndrome. *N Engl J Med* 2005; 352(16):1646-54.
- [206] Smith K, Leyden JJ. Safety of doxycycline and minocycline: A systematic review. *Clin Ther* 2005; 27(9): 1329-42.
- [207] Shapiro LE, Knowles SR, Shear NH. Comparative safety of tetracycline, minocycline, and doxycycline. *Arch Dermatol* 1997; 133(10): 1224-30.
- [208] Eriksson, A., Lepistö, M., Lundkvist, M., Munck, A.R.M., Zlatoidsky, P. Metalloproteinase inhibitors. US7427631 (2008) & US7666892 (2010)..
- [209] Tucker, H. Inhibitors of metalloproteinases. US6916817 (2005).
- [210] Henriksson, K., Henriksson, K.H. 2,5-Dioximidazolidin-4-yl acetamides and analogues as inhibitors of metalloproteinase MMP12. US7354940 (2010).
- [211] Fujimoto, R.A., McQuire, L.W., Monovich, L.G., Nantermet, P., Parker, D.T., Robinson, L.A., Skiles, J.W., Tommasi, R.A., Novartis, A.G. Azacycloalkyl substituted acetic acid derivatives for use as MMP inhibitors. WO2009072577 (2009).
- [212] Sawada, A., Neya, M. MMP Inhibitor. WO2002030873 (2002).
- [213] Pusateri, E.E., Tommasi, R.A., Leuter, T., Grob, J.E., Honda, A., Novartis, A.G. Selective hydroxamate based MMP inhibitors. WO2007117981 (2007).
- [214] Schwartz, M.A., Jin, Y., Sang, Q.X. Susbtituted heterocyclic mercaptosulfonamide metalloprotease inhibitors. WO2010028051 (2010).
- [215] McQuire, L.W., Rogel, O., Shultz, M., Tommasi, R.A., Weiler, S., Novartis, A.G. Selective hydroxamic acid based MMP-12 and MMP-13 inhibitors. WO2010007027 (2010).
- [216] Fryer, A.M., Rizzi, J. Alkylsulfonamide-substituted triazoles as matrix metalloprotease inhibitors. US20100234378 (2010).
- [217] Sheppeck, J.E., Duan, J., Xue, C.-B., Wasserman, Z. Hydantoins and related heterocycles as inhibitors of matrix metalloproteinases and/or TNF- α converting enzyme (TACE). US6906053 (2005).
- [218] Bird, J., Montana, J.G., Wills, R.E., Baxter, A.D., Owen, D.A. Selective MMP inhibitors having reduced side-effects. US6818622 (2004).
- [219] Wallach, S., Sameah-greenwald, S., Novik, A., Tsympkin, E., Nemzer, S. Novel nucleotide and amino acid sequences, and methods of use thereof for diagnosis. US20100261169 (2010).
- [220] Barta, T.E., Becker, D.P., Boehm, T.L., Decrescenzo, G.A., Willami, C.I., Mcdonald, J.J., Freskos, J.N., Getman, D.P., Hanson, G.J. Aromatic sulfone hydroxamic acid metalloprotease inhibitor. US20020039287 (2002).
- [221] Schotzinger, R.J. Metallo-hydrolase inhibitors using metal binding moieties in combination with targeting moieties. US20100256082 (2010).